



MONOCLONAL BLOOD GROUPING REAGENTS
DIRECTIONS FOR USE

Anti-P₁ Monoclonal: For Tube, DiaMed-ID and Ortho BioVue Techniques.

SUMMARY

Landsteiner discovered the P₁ antigen in 1927. Anti-P₁ does not generally react above room temperature and may often go undetected in routine testing. Anti-P₁ does not cause Haemolytic Disease of the Newborn and has only rarely been associated with Haemolytic Transfusion Reactions.

Anti-P ₁	Phenotype	Caucasians %	Afro-Americans %
+	P ₁	79	94
0	P ₂	21	6

PRINCIPLE

The reagent will cause agglutination (clumping) of test red cells, that carry the P₁ antigen, after centrifugation. No agglutination generally indicates the absence of the P₁ antigen (see **Limitations**).

REAGENT

Lorne Monoclonal IgM Anti-P₁ blood grouping reagent contains mouse monoclonal IgM antibodies prepared from the cell line, Clone 650, diluted in a solution containing sodium chloride and bovine albumin. The reagent is supplied at optimal dilution for use with all recommended techniques stated below without the need for further dilution or addition. For lot reference number and expiry date see **Vial Label**.

STORAGE

Reagent vials should be stored at 2 - 8°C on receipt. Prolonged storage at temperatures outside this range may result in accelerated loss of reagent reactivity. This reagent has undergone transportation stability studies at 37°C and -25°C as described in document EN13640:2002.

SAMPLE COLLECTION AND PREPARATION

Blood samples drawn with or without anticoagulant may be used for antigen typing. If testing is delayed, then store specimens at 2-8°C. EDTA and citrate samples should be typed within 7 days of collection. Samples collected into ACD, CPD or CPDA-1 may be tested up to 35 days from the date of withdrawal. All blood samples should be washed at least twice with PBS or Isotonic saline before being tested.

PRECAUTIONS

1. The reagent is intended for *in vitro* diagnostic use only.
2. If a reagent vial is cracked or leaking, discard the contents immediately.
3. Do not use the reagent past the expiration date (see **Vial Label**).
4. Do not use the reagent if a precipitate is present.
5. Protective clothing should be worn when handling the reagents, such as disposable gloves and a laboratory coat.
6. The reagent has been filtered through a 0.2 µm capsule to reduce the bio-burden. Once a vial has been opened the contents should remain viable up until the expiry date as long as there is no marked turbidity, which can indicate reagent deterioration or contamination.
7. The reagent contains <0.1% sodium azide. Sodium azide may be toxic if ingested and may react with lead and copper plumbing to form explosive metal azides. On disposal flush away with large volumes of water.
8. No known tests can guarantee that products derived from human or animal sources are free from infectious agents. Care must be taken in the use and disposal of each vial and its contents.

DISPOSAL OF REAGENT AND DEALING WITH SPILLAGES

For information on disposal of the reagent and decontamination of a spillage site see **Material Safety Data Sheets**, available on request.

CONTROLS AND ADVICE

1. It is recommended a positive control (ideally P₁ weak cells) and a negative control be tested in parallel with each batch of tests. Tests must be considered invalid if controls do not show expected results.
2. In the **Recommended Techniques** one volume is approximately 50µl when using the vial dropper provided.
3. The use of the reagent and the interpretation of results must be carried out by properly trained and qualified personnel in accordance with the requirements of the country where the reagent is in use.
4. The user must determine suitability of the reagent for use in other techniques.

REAGENTS AND MATERIALS REQUIRED

- DiaMed ID-Cards (Neutral).
- DiaMed ID-Centrifuge.
- DiaMed ID-CellStab).
- Glass test tubes (10 x 75 mm or 12 x 75 mm).
- Ortho BioVue System Cassettes (Neutral).
- Ortho BioVue System Centrifuge.
- Ortho 0.8% Red Cell Diluent.
- PBS solution (pH 6.8-7.2) or Isotonic saline solution (pH 6.5-7.5).
- Positive (ideally P₁ weak) and negative control red cells.
- Test tube centrifuge.
- Volumetric pipettes.

RECOMMENDED TECHNIQUES

A. Tube Technique

1. Prepare a 2-3% suspension of washed test red cells in PBS or Isotonic saline.
2. Place in a labelled test tube: 1 volume of Lorne reagent and 1 volume of test red cell suspension.
3. Mix thoroughly and incubate at 4°C ± 2°C for 15 minutes.
4. Centrifuge all tubes for 20 seconds at 1000 rcf or for a suitable alternative time and force.
5. Gently resuspend red cell button and read macroscopically for agglutination

B. DiaMed-ID Micro Typing Technique

1. Prepare a 0.8% suspension of washed test red cells in ID-CellStab.
2. Remove aluminium foil from as many microtubes as needed.
3. Place in appropriate microtube: 50µl of test red cell suspension and 25µl of Lorne reagent.
4. Incubate the ID-Card for 15 minutes at 4°C ± 2°C.
5. Centrifuge ID-Card in a DiaMed ID centrifuge.
6. Read macroscopically for agglutination.

C. Ortho BioVue Typing Technique

1. Prepare a 0.8% suspension of washed test red cells in 0.8% Ortho Red Cell Diluent.
2. Remove aluminium foil from as many reaction chambers as needed.
3. Place in appropriate reaction chamber: 50µl of test red cell suspension and 40µl of Lorne reagent.
4. Incubate the cassette(s) for 15 minutes at 4°C ± 2°C.
5. Centrifuge cassette(s) in an Ortho BioVue System Centrifuge.
6. Read macroscopically for agglutination.

INTERPRETATION OF TEST RESULTS

1. **Positive:** Agglutination of the test red cells constitutes a positive test result and within accepted limitations of test procedure, indicates the presence of the P₁ antigen on the test red cells.
2. **Negative:** No agglutination of the test red cells constitutes a negative result and within the accepted limitations of the test procedure, indicates the absence of the P₁ antigen on the test red cells.

STABILITY OF THE REACTIONS

1. Tests should be read immediately after centrifugation. Delays may result in dissociation of antigen-antibody complexes leading to false negative, or weak positive reactions.
2. Caution should be exercised in the interpretation of results of tests performed at temperatures other than those recommended.

LIMITATIONS

1. The P₁ antigen is poorly expressed on the cells of newborns.
2. There is a wide variation in the amount of P₁ antigen present on different P₁ positive cells. The strength of agglutination observed with such cells is likely to vary accordingly.
3. Stored blood may give weaker reactions than fresh blood.
4. False positive or false negative results may also occur due to:
 - Contamination of test materials
 - Improper storage, cell concentration, incubation time or temperature
 - Improper or excessive centrifugation
 - Deviation from the recommended techniques

SPECIFIC PERFORMANCE CHARACTERISTICS

1. The reagent has been characterised by all the procedures mentioned in the **Recommended Techniques**.
2. Prior to release, each lot of Lorne Monoclonal Anti-P₁ is tested by the **Recommended Techniques** against a panel of antigen-positive red cells to ensure suitable reactivity.
3. Specificity of source monoclonal antibody is demonstrated using a panel of antigen-negative cells.
4. The Quality Control of the reagent was performed using red cells that had been washed twice with PBS or Isotonic saline prior to use.
5. The reagent complies with the recommendations contained in the latest issue of the Guidelines for the UK Blood Transfusion Services.

DISCLAIMER

1. The user is responsible for the performance of the reagent by any method other than those mentioned in the **Recommended Techniques**.
2. Any deviations from the **Recommended Techniques** should be validated prior to use⁷.

BIBLIOGRAPHY

1. Kholer G, Milstein C. Continuous culture of fused cells secreting antibody of predefined specificity. Nature 1975; **256**, 495-497
2. Race RR, Sanger R. Blood Groups in Man, 6th Edition. Blackwell Scientific, Oxford 1975; Chapter 2.
3. Mollison PL. Blood Transfusion in Clinical Medicine, 8th Edition, Blackwell Scientific, Oxford 1987; Chapter 7.
4. Issitt PD. Applied Blood Group Serology, 3rd Edition. Montgomery Scientific, Miami 1985; Chapter 6
5. BSCH Blood Transfusion Task Force. Guidelines for microplate techniques in liquid-phase blood grouping and antibody screening, Clinical Laboratory Haematology 1990; **12**, 437-460.
6. Guidelines for the Blood Transfusion Service in the United Kingdom. H.M.S.O. Current Edition.
7. British Committee for Standards in Haematology, Blood Transfusion Task Force. Recommendations for evaluation, validation and implementation of new techniques for blood grouping, antibody screening and cross matching. Transfusion Medicine, 1995, **5**, 145-150.

AVAILABLE REAGENT SIZES








Vial Size	Catalogue Number
2 ml	315002
1000 ml	315000*

*This size is For Further Manufacturing Use (FFMU) only and is therefore not CE marked.

For the availability of other sizes, please contact:

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TABLE OF SYMBOLS

	Batch Number		<i>in-vitro</i> Diagnostic
	Catalogue Reference		Store At
	Expiry Date		Manufacturer
	Read Pack Insert		