MONOCLONAL BLOOD GROUPING REAGENTS
DIRECTIONS FOR USE

Anti-S and Anti-s Monoclonal: For Indirect Antiglobulin Techniques.

SUMMARY
The S and s antigens were reported in 1947 and 1951 respectively and form part of the MNS system. Anti-S and anti-s have both been implicated in Haemolytic Transfusion Reactions and Haemolytic Disease of the Newborn.

<table>
<thead>
<tr>
<th>Anti-S</th>
<th>Anti-s</th>
<th>Phenotype</th>
<th>Caucasians %</th>
<th>Afro-Americans %</th>
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<tbody>
<tr>
<td>+</td>
<td>-</td>
<td>S+</td>
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<tr>
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<td>S+</td>
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<tr>
<td>0</td>
<td>0</td>
<td>S-</td>
<td>0</td>
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PRINCIPLE
The reagents will cause indirect agglutination (clumping) of test red cells, that carry the corresponding specific antigen, in the antiglobulin phase of testing. No agglutination generally indicates the absence of the corresponding specific antigen (see Limitations).

REAGENTS
These Monoclonal IgG blood grouping reagents contain human monoclonal antibodies diluted in a phosphate buffer containing sodium chloride and bovine albumin. Each reagent is supplied at optimal dilution for use with all the recommended techniques stated below without the need for further dilution or addition. For lot number reference and expiry date see Vial Label.

STORAGE
Do not freeze. Reagent vials should be stored at 2–8ºC on receipt. Prolonged storage at temperatures outside this range may result in accelerated loss of reagent reactivity. This reagent has undergone transportation stability studies at 37ºC and −25ºC as described in document EN13640:2002.

SAMPLE COLLECTION AND PREPARATION
Blood samples drawn with or without anticoagulant may be used for antigen typing. If testing is delayed, then store specimens at 2–8ºC, EDTA and citrate samples should be typed within 7 days after collection. Samples collected into ACD, CPD or CPDA-1 may be tested up to 35 days from the date of withdrawal. All blood samples should be washed at least twice with PBS or Isotonic saline before being tested.

PRECAUTIONS
1. The reagents are intended for in vitro diagnostic use only.
2. If a reagent vial is cracked or leaking, discard the contents immediately.
3. Do not use the reagents past the expiration date (see Vial Label).
4. Do not use the reagents if a precipitate is present.
5. Protective clothing should be worn when handling the reagents, such as disposable gloves and a laboratory coat.
6. The reagents have been filtered through a 0.2 μm capsule to reduce the bio-burden. Once a vial has been opened the contents should remain viable up until the expiry date as long as there is no marked turbidity, which can indicate reagent deterioration or contamination.
7. The reagents contain 0.1% sodium azide. Sodium azide may be toxic if ingested and may react with lead and copper plumbing to form explosive metal azides. On disposal flush away with large volumes of water.
8. Materials used to produce the reagents were tested at source and found to be negative for HIV 1+2 and HCV antibodies and HBsAg using approved microbiological tests.
9. No known tests can guarantee that products derived from human or animal sources are free from infectious agents. Care must be taken in the use and disposal of each vial and its contents.

DISPOSAL OF REAGENT AND DEALING WITH SPILLAGES
For information on disposal of the reagents and decontamination of a spillage site see Material Safety Data Sheets, available on request.

CONTROLs AND ADVICE
1. It is recommended a positive control (ideally heterozygous cells) and a negative control be tested in parallel with each batch of tests. Tests must be considered invalid if controls do not show expected results.
2. The antiglobulin techniques can only be considered valid if all negative tests react positively with IgG sensitised red cells.
3. In the Tube Technique one volume is approximately 50μl when using the vial dropper provided.
4. The use of the reagents and the interpretation of results must be carried out by properly trained and qualified personnel in accordance with the requirements of the country where the reagents are in use.
5. The user must determine the suitability of the reagents for use in other techniques.

REAGENTS AND MATERIALS REQUIRED
- Anti-human globulin i.e. Lorne AHG Elite (Cat # 435010) or Anti-Human IgG i.e. Lorne Anti-Human IgG (Cat # 402010).
- Coombs cell washer.
- DiaMed ID-Cards (LISS/Coombs).
- DiaMed ID-Centrifuge.
- DiaMed ID-CellStab.
- DiaMed ID-Incubator equilibrated to 37°C ± 2°C.
- Glass test tubes (10 x 75 mm or 12 x 75 mm).
- IgG sensitised red cells i.e. Lorne Coombs Control Cells (Cat # 970010).
- Ortho BioVue System Cassettes (AHG/Coombs).
- Ortho BioVue System Centrifuge.
- Ortho BioVue System Heat Block equilibrated to 37°C ± 2°C.
- Ortho 0.8% Red Cell Diluent.
- PBS solution (pH 6.8–7.2) or Isotonic saline solution (pH 6.5–7.5).
- Positive (ideally heterozygous) and negative control red cells.
- Volumetric pipettes.
- Water bath or dry heat incubator equilibrated to 37°C ± 2°C.

RECOMMENDED TECHNIQUES
A. Indirect Antiglobulin Technique (IAT)
1. Prepare a 2-3% suspension of washed test red cells in PBS or Isotonic saline.
2. Place in a labelled test tube: 1 volume of Lorne reagent and 1 volume of test red cell suspension.
3. Mix thoroughly and incubate at 37°C for 15 minutes.
4. Washing test red cells 1 time with PBS or Isotonic saline, taking care to completely decant saline after the wash.
5. Add 2 volumes of anti-human globulin or anti-IgG to each dry cell button. Mix thoroughly and centrifuge all tubes for 20 seconds at 1000 rcf or for a suitable alternative time and force.
6. Gently resuspend red cell button and read macroscopically for agglutination.
7. Confirm validity of all negative reactions with IgG sensitised red cells.

B. DiaMed-ID Micro Typing Technique
1. Prepare a 0.8% suspension of washed test red cells in the ID-Diluent.
2. Remove aluminium foil from as many reaction chambers as needed.
3. Place in appropriate microtube: 50μl of test red cell suspension and 25μl of Lorne reagent.
4. Incubate the LISS/Coombs ID-Card(s) for 15 minutes at 37°C.
5. Centrifuge the LISS/Coombs ID-Card(s) in the DiaMed ID-Centrifuge.
6. Read macroscopically for agglutination.

C. Ortho BioVue Typing Technique
1. Prepare a 0.8% suspension of washed test red cells in 0.8% Ortho Red Cell Diluent.
2. Remove aluminium foil from as many reaction chambers as needed.
3. Place in appropriate reaction chamber: 50μl of test red cell suspension and 40μl of Lorne reagent.
4. Incubate the cassette(s) for 15 minutes at 37°C.
5. Centrifuge cassette(s) for 5 minutes in an Ortho BioVue System Centrifuge.
6. Read macroscopically for agglutination.

INTERPRETATION OF TEST RESULTS
1. Positive: Agglutination of the test red cells constitutes a positive test result and within accepted limitations of test procedure, indicates the presence of the appropriate antigen on the test red cells.
2. Negative: No agglutination of the test red cells constitutes a negative result and within the accepted limitations of the test procedure, indicates the absence of the appropriate antigen on the test red cells.

STABILITY OF THE REACTIONS
1. Washing steps should be completed without interruption and tests centrifuged and read immediately after addition of the reagent. Delays may result in dissociation of antigen-antibody complexes, causing false negative or weak positive results.
2. Caution should be exercised in the interpretation of results of tests performed at temperatures other than those recommended.
LIMITATIONS
1. Red cells that have a positive DAT due to a coating of IgG cannot be typed by the Indirect Antiglobulin Technique.
2. Suppressed or diminished expression of certain blood group antigens may conversely give rise to false negative reactions and so caution should always be exercised when assigning genotypes on the basis of test results.
3. False positive or false negative results may also occur due to:
   - Contamination of test materials
   - Improper storage, cell concentration, incubation time or temperature
   - Improper or excessive centrifugation
   - Deviation from the recommended techniques

SPECIFIC PERFORMANCE CHARACTERISTICS
1. The reagents have been validated by the procedures mentioned in the Recommended Techniques.
2. Prior to release, each lot of Lorne Anti-S and Anti-s reagent is tested by the Recommended Techniques against a panel of antigen-positive red cells to ensure suitable reactivity.
3. Specificity of source monoclonal antibodies is demonstrated using a panel of antigen-negative cells
4. The Quality Control of the reagents was performed using red cells that had been washed twice with PBS or Isotonic saline prior to use.
5. The reagents comply with the recommendations contained in the latest issue of the Guidelines for the UK Blood Transfusion Services in the United Kingdom.

DISCLAIMER
1. The user is responsible for the performance of the reagents by any method other than those mentioned in the Recommended Techniques.
2. Any deviations from the Recommended Techniques should be validated prior to use.

BIBLIOGRAPHY

AVAILABLE REAGENT SIZES

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*This size is For Further Manufacturing Use (FFMU) only is therefore not CE marked.

For the availability of other sizes, please contact:

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