



MONOCLONAL BLOOD GROUPING REAGENTS
DIRECTIONS FOR USE

Anti-Fy^a Monoclonal: For Indirect Antiglobulin Techniques.

SUMMARY

The Fy^a and Fy^b antigens were reported in 1950 and 1951 respectively. Anti-Fy^a and anti-Fy^b have both been implicated in immediate and delayed Haemolytic Transfusion Reactions and Haemolytic Disease of the Newborn.

Anti-Fy ^a	Anti-Fy ^b	Phenotype	Caucasians %	Afro-Americans %
+	0	Fy(a+b-)	17	9
0	+	Fy(a-b+)	34	22
+	+	Fy(a+b+)	49	1
0	0	Fy(a-b-)	Very Rare	68

INTENDED PURPOSE

The reagent is a blood grouping reagent intended to be used to qualitatively determine the presence or absence of the Fy(a) antigen (FY001) on the red cells of blood donors or patients requiring a blood transfusion when tested in accordance with the recommended techniques stated in this IFU.

PRINCIPLE

The reagent will cause indirect agglutination (clumping) of red cells, that carry the Fy(a) antigen, in the antiglobulin phase of testing. No agglutination generally indicates the absence of the Fy(a) antigen (see **Limitations**).

REAGENTS

This Monoclonal IgG blood grouping reagent contains human monoclonal antibodies diluted in a phosphate buffer containing sodium chloride and bovine albumin. The reagent does not contain or consist of CMR substances, or endocrine disrupting substances or that could result in sensitisation or an allergic reaction by the user. The reagent is supplied at optimal dilution for use with all the recommended techniques stated below without the need for further dilution or addition. For lot reference number and expiry date see **Vial Label**.

Product	Cell Line/Clone
Anti-Fya	DG-FYA-02

STORAGE

Do not freeze. Reagent vials should be stored at 2 - 8°C on receipt. Prolonged storage at temperatures outside this range may result in accelerated loss of reagent reactivity. This reagent has undergone transportation stability studies at 37°C and -25°C as described in document BS EN ISO 23640:2015.

SAMPLE COLLECTION AND PREPARATION

Blood samples can be collected into EDTA, citrate, CPDA anticoagulants or as a clotted sample. The samples should be tested as soon as possible following collection. If a delay in testing should occur, store the samples at 2-8°C. Samples displaying gross haemolysis or microbial contamination should not be used for testing. Blood samples showing evidence of lysis may give unreliable results. It is preferable (but not essential) to wash all blood samples with PBS or Isotonic saline before being tested.

PRECAUTIONS

- The reagents are intended for *in vitro* diagnostic use only.
- If a reagent vial is cracked or leaking, discard the contents immediately.
- Do not use the reagents past the expiration date (see **Vial Label**).
- Do not use the reagents if a precipitate is present.
- Protective clothing should be worn when handling the reagents, such as disposable gloves and a laboratory coat.
- The reagents have been filtered through a 0.2 µm capsule to reduce the bio-burden, but is not supplied sterile. Once a vial has been opened the contents should remain viable up until the expiry date as long as there is no marked turbidity, which can indicate reagent deterioration or contamination.
- The reagents contain < 0.1% sodium azide. Sodium azide may be toxic if ingested and may react with lead and copper plumbing to form explosive metal azides. On disposal flush away with large volumes of water.
- Materials used to produce the reagents were tested at source and found to be negative for HIV 1+2 and HCV antibodies and HBsAg using approved microbiological tests.
- No known tests can guarantee that products derived from human or animal sources are free from infectious agents. Care must be taken in the use and disposal of each vial and its contents.

DISPOSAL OF REAGENT AND DEALING WITH SPILLAGES

For information on disposal of the reagents and decontamination of a spillage site see **Material Safety Data Sheets**, available on request.

CONTROLS AND ADVICE

- It is recommended a positive control (ideally heterozygous cells) and a negative control be tested in parallel with each batch of tests. Tests must be considered invalid if controls do not show expected results.
- The antiglobulin techniques can only be considered valid if all negative tests react positively with IgG sensitised red cells.
- Before use, let the reagent warm up to room temperature. As soon as the reagent has been used, put the reagent back in storage at 2-8°C.
- In the **Tube Technique** one volume is approximately 50µl when using the vial dropper provided.
- The use of the reagents and the interpretation of results must be carried out by properly trained and qualified personnel in accordance with the requirements of the country where the reagents are in use.
- User must determine suitability of the reagents for use in other techniques.

REAGENTS AND MATERIALS REQUIRED

- Anti-human globulin i.e. Lorne AHG Elite (Cat # 435010 or 415010) or Anti-Human IgG i.e. Lorne Anti-Human IgG (Cat # 402010 or 401010).
- Coombs cell washer.
- Bio-Rad ID-Cards (LISS/Coombs or Coombs Anti-IgG).
- Bio-Rad ID-Centrifuge.
- Bio-Rad ID-CellStab or ID-Diluent 2.
- Bio-Rad ID-Incubator equilibrated to 37°C ± 2°C.
- Glass test tubes (10 x 75 mm or 12 x 75 mm).
- IgG sensitised red cells i.e. Lorne Coombs Control Cells (Cat # 970010).
- Ortho BioVue System Cassettes (AHG Polyspecific or AHG Anti-IgG).
- Ortho BioVue System Centrifuge.
- Ortho BioVue System Heat Block equilibrated to 37°C ± 2°C.
- Ortho 0.8% Red Cell Diluent.
- PBS solution (pH 6.8-7.2) or Isotonic saline solution (pH 6.5-7.5).
- Positive (ideally heterozygous) and negative control red cells.
- Volumetric pipettes.
- Water bath or dry heat incubator equilibrated to 37°C ± 2°C.

RECOMMENDED TECHNIQUES

A. Indirect Antiglobulin Technique (IAT)

- Prepare a 2-3% suspension of red cells in PBS or Isotonic saline.
- Place in a labelled test tube: 1 volume of Lorne reagent and 1 volume of red cell suspension.
- Mix thoroughly and incubate at 37°C for 15 minutes.
- Wash red cells 1 time with PBS or Isotonic saline, taking care to decant saline after the wash.
- Add 2 volumes of anti-human globulin or anti-IgG to each dry cell button.
- Mix thoroughly and centrifuge all tubes for 20 seconds at 1000 rcf or for a suitable alternative time and force.
- Gently resuspend red cell button and read macroscopically for agglutination
- Confirm validity of all negative reactions with IgG sensitised red cells.

B. Bio-Rad ID Technique (LISS/Coombs cards)

- Prepare a 0.8% suspension of red cells in ID-CellStab or ID-Diluent 2.
- Remove aluminium foil from as many microtubes on either LISS/Coombs or Coombs Anti-IgG ID-Card(s) as needed.
- Place in appropriate microtube: 50µl of red cell suspension and 25µl of Lorne reagent.
- Incubate the LISS/Coombs ID-Card(s) for 15 minutes at 37°C.
- Centrifuge the LISS/Coombs ID-Card(s) in the Bio-Rad ID-Card centrifuge.
- Read macroscopically for agglutination.

C. Ortho BioVue Technique (AHG cassettes)

- Prepare a 0.8% suspension of red cells in 0.8% Ortho Red Cell Diluent.
- Remove aluminium foil from as many reaction chambers on either AHG Polyspecific or AHG Anti-IgG cassettes as needed.
- Place in appropriate reaction chamber: 50µl of red cell suspension and 40µl of Lorne reagent.
- Incubate the cassette(s) for 15 minutes at 37°C.
- Centrifuge cassette(s) in an Ortho BioVue System Centrifuge.
- Read macroscopically for agglutination.

INTERPRETATION OF TEST RESULTS

- Positive:** Agglutination of the red cells constitutes a positive test result and within accepted limitations of test procedure, indicates the presence of the appropriate antigen on the red cells.
- Negative:** No agglutination of the red cells constitutes a negative result and within the accepted limitations of the test procedure, indicates the absence of the appropriate antigen on the red cells.

STABILITY OF THE REACTIONS

1. Washing steps should be completed without interruption and tests centrifuged and read immediately after addition of the reagent. Delays may result in dissociation of antigen-antibody complexes, causing false negative or weak positive results.
2. Caution should be exercised in the interpretation of results of tests performed at temperatures other than those **recommended**.

LIMITATIONS

1. Red cells that have a positive DAT due to a coating of IgG cannot be typed by the **Indirect Antiglobulin Technique**.
2. Suppressed or diminished expression of certain blood group antigens may conversely give rise to false negative reactions and so caution should always be exercised when assigning genotypes on the basis of test results.
3. False positive or false negative results may also occur due to:
 - Contamination of test materials
 - Improper storage, cell concentration, incubation time or temperature
 - Improper or excessive centrifugation
 - Deviation from the recommended techniques

SPECIFIC PERFORMANCE CHARACTERISTICS

1. Prior to release, each lot of reagent was tested using the recommended test methods listed in this IFU. The tests complied with the test requirements as stated in the current version/issue of the "Guidelines for the Blood Transfusion Services in the United Kingdom".
2. Specificity of source monoclonal antibodies is demonstrated using a panel of antigen-negative cells
3. The Quality Control of the reagents was performed using red cells with phenotypes that were verified by a UK blood transfusion centre and had been washed with PBS or Isotonic saline prior to use.

DISCLAIMER

1. The user is responsible for the performance of the reagents by any method other than those mentioned in the **Recommended Techniques**.
2. Any deviations from the **Recommended Techniques** should be validated prior to use⁶.

BIBLIOGRAPHY

1. Widman FK. Technical Manual, 9th Edition. American Association of Blood Banks, Arlington, VA, 1985; Chapter 8
2. Race RR, Sanger R. Blood Groups in Man, 6th Edition. Blackwell Scientific, Oxford 1975; Chapter 2
3. Mollison PL. Blood Transfusion in Clinical Medicine, 8th Edition. Blackwell Scientific, Oxford 1987; Chapter 7
4. Issitt PD. Applied Blood Group Serology, 3rd Edition. Montgomery Scientific, Miami 1985; Chapter 6
5. Guidelines for the Blood Transfusion Service in the United Kingdom. H.M.S.O. Current Edition.
6. British Committee for Standards in Haematology, Blood Transfusion Task Force. Recommendations for evaluation, validation and implementation of new techniques for blood grouping, antibody screening and cross matching. Transfusion Medicine, 1995, 5, 145-150.

AVAILABLE REAGENT SIZES

	Vial Size	Catalogue Number	Tests per vial
Anti-Fya	2 ml	774002	40
Monoclonal	1000 ml	774000*	20,000

*This size is For Further Manufacturing Use (FFMU) only and is therefore not CE marked.



Lorne Laboratories Limited
 Unit 1 Cutbush Park Industrial Estate
 Danehill
 Lower Earley
 Berkshire, RG6 4UT
 United Kingdom
 Tel: +44 (0) 118 921 2264
 Fax: +44 (0) 118 986 4518
 E-mail: info@lornelabs.com

EC	REP	Advena Ltd. Tower Business Centre, 2 nd Fl., Tower Street, Swatar, BKR 4013, Malta
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